Anti-FMC63 mAb (APC)

PRODUCT INFORMATION

Catalog Number	GTU-ADA-FMC63-Ab01-APC200
Product Name	Anti-FMC63 mAb (APC)
Size	200 T
Source	Anti-FMC63 mAb (APC) antibody was coupled with allophycocyanin (APC) fluorescent dye under optimal conditions. The reactivity of this antibody conjugate is expected to be similar to that of the uncoupled Anti-FMC63 mAb [GTU-ADA-FMC63-Ab-APC200], which is expected to react with FMC63 scFv on the cell surface.
Isotype	Mouse IgG2a
Immunogen	scFv region of a CD19-specific mouse mAb clone FMC63
Specificity/Sensitivity	Recognizes FMC63 scFv specifically
Species reactivity	Any species and cells expressing FMC63 scFv
Storage temperature	2-8° C
Conjugate	APC
Applications	FACS Analysis 5 μL/test, in 100-200 μL with~1×10^6 cells
Formulation	Phosphate-buffered solution (pH 7.2), containing 0.03% Proclin 300 and 5 mg/mL BSA
Quality Control Data	Molar APC /Ab Ratio 1.07 Final Volume 1 mL 0.2 μm Filtration Yes

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SHIPPING & STORAGE

Fluorescent-labeled antibodies are shipped in an amber vial and should be stored at 4°C protected from exposure to light.

Do not expose the fluorescent-conjugated antibody to light. Continuous exposure to light will cause the antibody to gradually lose its fluorescence.

DESCRIPTION

FMC63 is a CD19-specific mouse monoclonal antibody, which is a target for immunotherapy of B-lineage leukemia and lymphoma. FMC63 scFv is the most commonly used extracellular domain component of CD19-specific CARs. Most of the reported CART-CD19 clinical trials contain anti-CD19 scFv derived from FMC63, such as the two FDA-approved CARs Kymriah and Yescarta. This product was verified to specifically recognize the antigen recognition domain of FMC63-derived CAR.

Flow Cytometry (FACS) Protocols

Sample preparation

• Typically use 1-2×10^6 cells in a 100-200 µL experimental sample (per test). Since applications vary, each investigator should titrate the reagent to obtain optimal results.

• Cells should be rinsed with PBS to remove serum proteins prior to antibody staining. If staining with more than one antibody, prepare a pool of antibodies together.

• All incubations should be performed with minimal light exposure.

Antibody Staining Protocol

1. Add 5 µL of labeled appropriate antibody directly to the 100-200 µL of rinsed cells.

2. Incubate at Room Temperature or 4°C and away from light for 30 minutes.

3. Add 1 mL PBS to rinse non-bound antibody. Centrifuge at 1200-1500 rpm for 5 minutes, decant the supernatant from the cell pellet.

4. Rinse once again.

5. Add 200-400 µL of PBS to resuspend the cells.

6. The sample is ready for Flow analysis.

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Note: Fc Block

1. Samples of cells without Fc Receptors can skip the Fc Block step.

2. Phagocytic cells (such as macrophages and granulocytes) can bind non-specifically to antibodies unless the Fc receptors of these cells are blocked. If the sample is from a tissue homogenization possibly containing macrophages, or from a cell culture line expressing Fc receptors (such as Daudi and THP-1), you can use anti-Fc Receptor nonlabelled antibody (anti-CD16/32 in mice or human IgG in human) to block the receptors.

3. Add 1 µg of Fc Block Antibody directly or 5 µg human IgG to 100 µL of suspended cells.

4. Incubate on ice for 20 minutes.

5. Add 1 mL PBS to rinse non-bound antibody. Centrifuge at 1200-1500 rpm for 5 minutes, decant the supernatant from the cell pellet.

6. Rinse once again.

7. Add 100-200 µL of PBS to resuspend the cells.

8. Proceed directly to Antibody Staining Protocol.

Conditions:
The information disclosed herein is not to be construed as a recommendation to use the above product in violation of any patents.
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