

PurProX™ AAVEasy rapid-to-purify Spin Column

Product Instruction Manual

Product Information

Classification	Description
Catalog number	GMV-PurProX-AAVEasy-U-1
Product name	PurProX™ AAVEasy Spin Column

Buffer Configuration

Binding buffer: 1*PBS + 0.35M NaCl + 0.01% F-68, pH7.4

• Elution buffer: 0.1 M Glycine + 2mM MgCl₂ + 0.01%F-68, pH2.5

Stripping buffer: 0.1 M phosphoric acid + 0.01% F-68, pH2.0

Neutralization buffer: 1M Tris

Note:

- 1. Entire purification process was performed at room temperature;
- 2. Do not tighten the centrifuge tube cap;
- 3. If the filler "flies off" during the purification process, it will not affect the final purification, and you can continue the experimental operation.

Protocol

- 1. Sample processing
- AAV crude extract (cell lysate after omnipotent enzyme treatment) was centrifuged at 8000 rpm for 10 minutes at room temperature;
- 2) Take the AAV supernatant after centrifugation and filter it with a $0.22\,\mu$ m filter to obtain the AAV to be purified;
- 2. Column balance

Add 5 mL of Binding buffer to the prepared purification column and centrifuge at 100*g for 1 min, Repeat 3 times;

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Loading

Place the treated AAV to be purified (recommended to dilute to 10 ml with Binding buffer) into the assembled purification column, centrifuge at 100*g for 1 minute at room temperature, then add the liquid in the centrifuge tube back into the purification column, repeat 3 times, and remove the liquid in the centrifuge tube;

4. Cleaning

Add 10 mL of Binding buffer to the purification column, centrifuge at 100*g for 2 minutes, remove the liquid in the centrifuge tube, and repeat 5 times;

- 5. Elution
- 1) After cleaning, place in an airtight container at 200*g for 1 minute;
- 2) Transfer the purification column to a new centrifuge tube, add 0.5 mL of Elution buffer, centrifuge at 100*g for 1 min to elute, do not discard the liquid, then add 0.5 mL of Elution buffer, centrifuge at 100*g for 1 min to obtain the eluate;
- 3) Quickly transfer the eluate to a 1.5 mL centrifuge tube containing 80 $\,\mu$ L of neutralization buffer and mix thoroughly;

Note: After neutralization with neutralization buffer, pH test paper can be used to check whether the pH is neutral. Elution buffer or neutralization buffer can be used for fine-tuning.

- 6. Column regeneration
- Add 10 mL of Stripping buffer to the purification column, centrifuge at 100*g for 2 minutes, discard the liquid, and repeat 3 times;
- 2) Add 10 mL of Binding buffer, centrifuge at 100*g for 2 minutes, discard the liquid, and repeat 2 times:
- 7. Column preservation
- 1) Add 10 mL of 20% ethanol to the purification column, centrifuge at 100*g for 1 min, discard the liquid, and repeat 2 times;
- 2) Fill the internal sample tube with 20% ethanol and tighten the lower plug. Storage conditions: 20% ethanol, 4° C.

Note: After using the product, make sure the filler is completely soaked in 20% ethanol before storage.

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Storage

Storage conditions: $2 - 8^{\circ}$ C.

Shelf life: At least 6 months (6-month stability testing has been completed; long-term stability is being evaluated).







