

# PurProX™ AAVEasy rapid-to-purify Spin Column

## Product Instruction Manual

### Product Information

| Classification | Description                  |
|----------------|------------------------------|
| Catalog number | GMV-PurProX-AAVEasy-U-1      |
| Product name   | PurProX™ AAVEasy Spin Column |

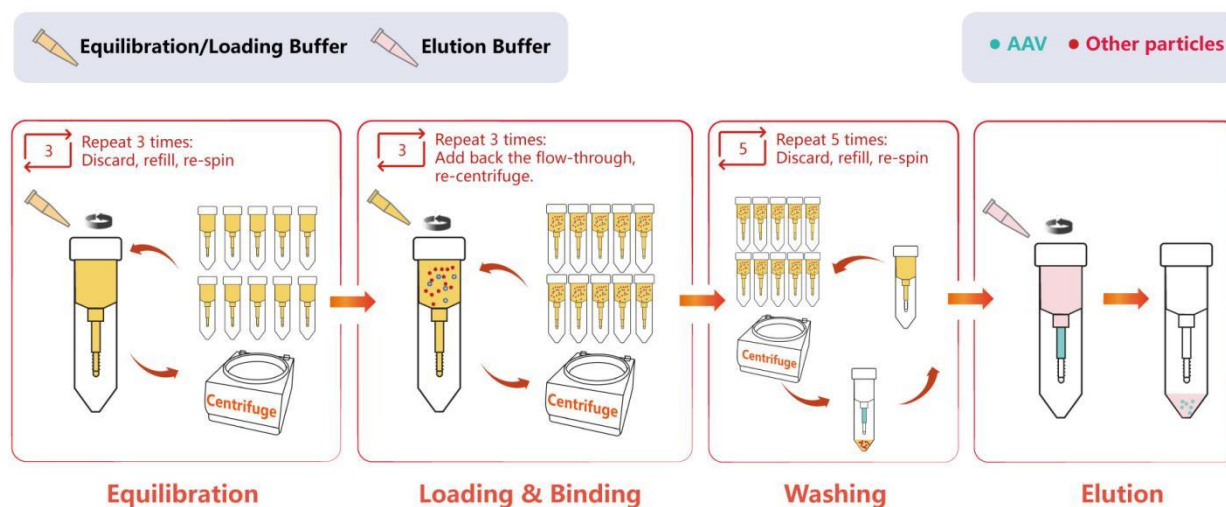
### Buffer Configuration

- Binding buffer: 1×PBS + 0.35M NaCl + 1×GMXbuffer01, pH7.4
- Elution buffer: 0.1 M Glycine + 2mM MgCl<sub>2</sub> + 1×GMXbuffer01, pH2.5
- Stripping buffer: 0.1 M phosphoric acid + 1×GMXbuffer01, pH2.0
- Neutralization buffer: 1M Tris

Note:

- 1、GeneMedi provides 1000×GMXbuffer01.
- 2、Entire purification process was performed at room temperature;
- 3、Do not tighten the centrifuge tube cap;
- 4、If the filler "flies off" during the purification process, it will not affect the final purification, and you can continue the experimental operation.

### PurProX® AAVEasy Purification Process



## Protocol

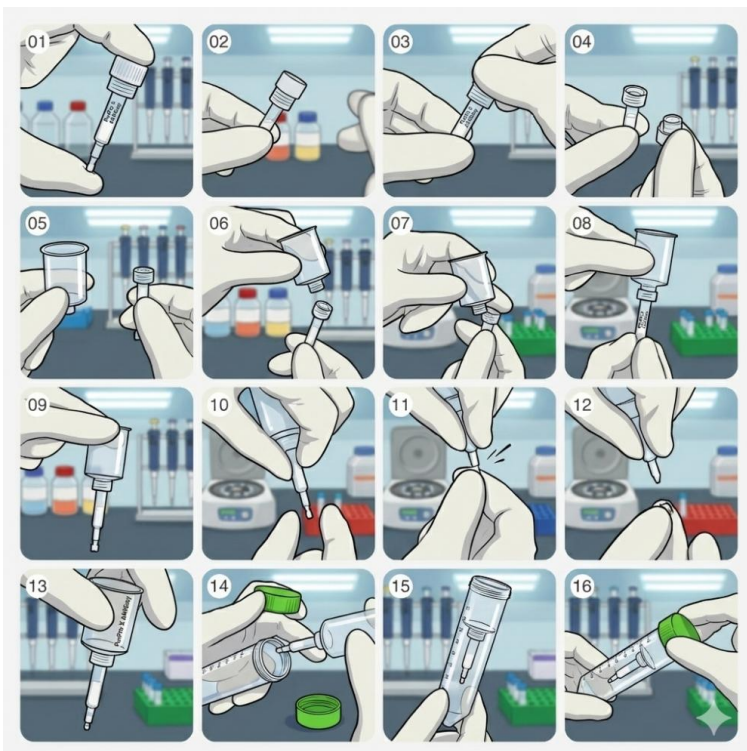
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### 1. Sample processing

- 1) AAV crude extract (cell lysate after omnipotent enzyme treatment) was centrifuged at 8000 rpm for 10 minutes at room temperature;
- 2) Take the AAV supernatant after centrifugation and filter it with a 0.22  $\mu$  m filter to obtain the AAV to be purified;

### 2. Column balance

To install the column, please follow the steps below:



Add 5 mL of Binding buffer to the prepared purification column and centrifuge at 100\*g for 1 min, Repeat 3 times;

### 3. Loading

Place the treated AAV to be purified (recommended to dilute to 10 ml with Binding buffer) into the assembled purification column, centrifuge at 100\*g for 1 minute at room temperature, then add the liquid in the centrifuge tube back into the purification column, repeat 3 times, and remove the liquid in the centrifuge tube;

#### 4. Cleaning

Add 10 mL of Binding buffer to the purification column, centrifuge at 100\*g for 2 minutes, remove the liquid in the centrifuge tube, and repeat 5 times;

#### 5. Elution

- 1) After cleaning, place in an airtight container at 200\*g for 1 minute;
- 2) Transfer the purification column to a new centrifuge tube, add 0.5 mL of Elution buffer, centrifuge at 100\*g for 1 min to elute, do not discard the liquid, then add 0.5 mL of Elution buffer, centrifuge at 100\*g for 1 min to obtain the eluate;
- 3) Quickly transfer the eluate to a 1.5 mL centrifuge tube containing 80  $\mu$  L of neutralization buffer and mix thoroughly;

Note: After neutralization with neutralization buffer, pH test paper can be used to check whether the pH is neutral. Elution buffer or neutralization buffer can be used for fine-tuning.

#### 6. Column regeneration

- 1) Add 10 mL of Stripping buffer to the purification column, centrifuge at 100\*g for 2 minutes, discard the liquid, and repeat 3 times;
- 2) Add 10 mL of Binding buffer, centrifuge at 100\*g for 2 minutes, discard the liquid, and repeat 2 times;

#### 7. Column preservation

- 1) Add 10 mL of 2% Phenylmethanol to the purification column, centrifuge at 100\*g for 1 min, discard the liquid, and repeat 2 times;
- 2) Fill the internal sample tube with 2% Phenylmethanol and tighten the lower plug. Storage conditions: 2% Phenylmethanol, 4° C.

Note: After using the product, make sure the filler is completely soaked in 2% Phenylmethanol before storage.

## Storage

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**Storage conditions:** 2 – 8° C.

**Shelf life:** At least 6 months (6-month stability testing has been completed; long-term stability is being evaluated).